Articles

Solid-Phase Synthesis of the Alkenyldiarylmethane (ADAM) Series of Non-Nucleoside HIV-1 Reverse Transcriptase Inhibitors

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Received January 10, 2001

The Sonogashira and Stille cross-coupling reactions have been employed in the synthesis of several non-nucleoside reverse transcriptase inhibitors (NNRTIs) in the alkenyldiarylmethane (ADAM) series. The synthesis has been carried out both in solution and on a solid support. In contrast to previous syntheses of NNRTIs in the ADAM series, the present strategy allows the incorporation of differently substituted aromatic rings in a stereochemically defined fashion. The most potent of the new ADAMs inhibited the cytopathic effect of HIV- 1_{RF} in CEM-SS cell culture with an EC $_{50}$ value of 20 nM.

The alkenyldiarylmethanes (ADAMs) are a class of HIV-1 non-nucleoside reverse transcriptase inhibitors (NNRTIs). 1-5 Certain ADAMs (e.g., ADAM 1) have been found to inhibit of the cytopathic effect of HIV-1 in cell culture at low nanomolar concentrations.^{4,5} A number of HIV-1 strains containing AZT resistance mutations have shown increased sensitivity to some of the ADAMs. indicating a possible therapeutic role for the ADAMs in combination with AZT.⁴ In general, the clinical use of NNRTIs in combination with two additional anti-HIV agents decreases HIV-1 RNA levels, increases CD4 lymphocyte counts, and delays disease progression. However, several factors continue to limit the selection of NNRTIs for combination chemotherapy, including drug incompatibilities, adverse effects, the emergence of resistant viral strains, and cross-resistance. $^{6-10}$ Therefore,

a need exists for additional NNRTIs that might display novel resistance mutation profiles, more favorable pharmacokinetic properties, and limited toxicities. We have therefore continued to design and synthesize additional NNRTIs in the ADAM series.

The previously existing ADAMs (e.g., 1) have been synthesized by attaching the alkenyl chain to a symmetrical benzophenone (e.g., 2) by Wittig, 1-4 McMurry, 5 or Horner-Emmons⁴ reactions. Since these methods proceed from the readily available, symmetrical benzophenones, they are not ideal for preparing ADAMs having nonidentical aromatic substituents. In addition, even if unsymmetrical aromatic benzophenones were used, these methods would be complicated by generation of cis and trans double bond isomers that would have to be separated and defined structurally. We have therefore sought to establish alternative syntheses of the ADAMs that would allow for the construction of compounds having nonidentical aromatic substituents with stereochemically defined relationships to the alkenyl side chain. Another goal of the present research project was to adapt the resulting solution-phase synthesis to the solid phase, which could eventually allow automation and the preparation of combinatorial libraries for lead optimization. 11-13

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The application of palladium-catalyzed reactions for C-C bond formation is particularly attractive for solidphase combinatorial library syntheses since the conditions employed are mild and many functional groups are compatible. 14-16 The present article describes the application of the Sonogashira and Stille cross-coupling reactions for the solution and solid-phase synthesis of stereochemically defined ADAMs having differently substituted aromatic rings. As shown by the retrosynthetic analysis described in Scheme 1, Stille coupling of 4 and 5 (X and Y = I or $SnBu_3$) would be expected to result in the ADAM **3**. The coupling partner **5** (in the case of $Y = SnBu_3$) could in turn be made by the Sonogashira reaction of the alkyne 6 and the aryl iodide 7, followed by hydrostannation. Since the Stille reaction proceeds with retention of the stereochemical integrity of the coupling partners, and hydrostannation of alkynes results in regiochemically defined cis addition to the triple bond, the stereochemistry (cis vs trans) of the side chain relative to the two aromatic rings in the final product would be predictable (i.e., the side chain would be cis to the aromatic ring containing R^2 in structure 3).

Results and Discussion

As shown in Scheme 2, 3-methylsalicylic acid (8) was quantitatively converted into its methyl ester using

^a Reagents and conditions: (a) TMSCHN₂/MeOH, benzene, 23 °C (2 h); (b) NaI, NaOH/MeOH, 0-3 °C (3 h) then NaOCl (5.25%); (c) dimethyl sulfate, K2CO3, acetone, reflux (24 h); (d) Bu3Sn-SnBu₃, Pd(PPh₃)₄, Pd(OAc)₂, triethylamine, 95-100 °C (3 h); (e) $\mathit{N}\text{-methyl}$ 5-hexynoamide, $(Ph_3P)_2\check{P}d(OAc)_2$, CuI, triethylamine, EtOAc, 23 °C (14 h); (f) Bu₃SnH, Pd(PPh₃)₄, THF, 23 °C (14 h); (g) I_2 , CH_2Cl_2 , 23 °C (20 min).

TMSCHN₂ in a mixture of MeOH and benzene.¹⁷ The methyl ester was then iodinated with sodium iodide in the presence of sodium hypochlorite as an oxidant.¹⁸ Treatment of the iodinated ester **9** with dimethyl sulfate and potassium carbonate in refluxing acetone produced methyl 5-iodo-3-methyl-2-methoxybenzoate 10 in 90% overall yield. The tributyltin derivative 11 was synthesized by heating a mixture of iodo ester 10, bis(tributyltin), tetrakis(triphenylphosphine)palladium, and palladium acetate in triethylamine at 95-100 °C overnight.18 The Sonogashira reaction of N-methyl 5-hexynoamide with iodo ester 10 afforded alkyne 12, 19,20 which underwent hydrostannation in the presence of Pd(PPh₃)₄ to afford the regio- and stereodefined vinylstannane 13 in 87% yield.^{21,22} Subsequent treatment of 13 with I₂ in CH₂Cl₂ produced vinyl iodide 14.²³

Our initial attempts to prepare ADAM 15 (Scheme 3), by employing vinyl tributylstanne 13 and aryl iodide 10 (or the alternative combination, vinyl iodide ${\bf 14}$ and aryl

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Scheme 3

13 or 14 (Y = SnBu3 or I)

tributylstanne 11) in the presence of Pd(PPh₃)₄, were unsuccessful because of the low yield of the coupling reaction.²⁴ In general, densely substituted stannanes reacts poorly, and their coupling must be carefully optimized. Considering the Stille coupling catalytic cycle, a better combination of catalyst, ligand, additive, and solvent might be beneficial to the cross-coupling reaction.²⁵ It has been shown that ligands with donicity toward Pd(II) lower than that of PPh3 (i.e., AsPh3) can lead to major (up to 1000-fold) rate enhancement in the transmetalation. Besides the ligand, the addition of fluoride might lead to more efficient coupling, perhaps by facilitating transmetalation from tin to palladium, since tin is fluorophilic.^{26–28} The cross-coupling reactions of organosilicon compounds are known to be facilitated by fluoride.^{29–32} Under the "optimized" conditions (Pd₂-(dba)₃/AsPh₃/CsF/DMF), Stille cross-coupling of vinyl tributylstannane 13 and aryl ioide 10 (or vinyl iodide 14 and aryl tributylstannane 11) afforded 15 in 74% (or 80%) isolated yield. The reactions proceeded exclusively with the desired ipso substitution, and none of the undesired cine substitution products were detected in the reaction mixtures.

 a Reactions and conditions: (a) Dimethyl sulfate, $K_2CO_3,$ acetone, reflux (24 h); (b) $SO_2Cl_2,$ $CH_2Cl_2,$ $50\,^\circ C$ (14 h); (c) aq. NaOH/MeOH, 60 $^\circ C$ (3 h); (d) DCC, HOBt/DMF, 23 $^\circ C$ (24 h); (e) methyl 5-hexynoate, Pd(OAc)2, CuI, Et_3N, DMF, 23 $^\circ C$ (12 h); (f) Bu_3SnH, Pd(PPh_3)4, THF, 23 $^\circ C$ (4 h); (g) Et_3N, MeOH (15:85, v/v), 50 $^\circ C$ (16 h); (h) 10 or 17, Pd2(dba)3, AsPh3, CsF/DMF, 80 $^\circ C$ (12 h); (i) NaOMe, MeOH, THF (1:2), reflux (20 h).

Having explored the performance of the above reaction sequence in solution, we went on to apply it on a solid support. The building block **19** was prepared through a three-step synthesis (Scheme 4). Methylation of both the

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compd	RT IC $_{50}$ rCdG a (μ M)	$ ext{EC}_{50} ext{MTS assay}^b \ (\mu ext{M})$	CC ₅₀ MTS assay ^c (μ M)	TI^d
AZT		0.005	>1.0	>201
15	0.804	2.26	17.10	7
26	0.516	0.020	1.46	73
27	0.587	0.080	2.17	27
28	0.300	0.013	31.6	2430

^a Inhibitory activity vs HIV-1 reverse transcriptase with rCdG as the template:primer. ^b The EC₅₀ is the 50% inhibitory concentration for cytopathicity of HIV-1_{RF} in CEM-SS cells. ^c The CC₅₀ is the 50% cytotoxic concentration for mock-infected CEM cells. ^d The TI is the therapeutical index, which is the CC₅₀ divided by the EC₅₀.

phenol and carboxylic acid groups of 16 using dimethyl sulfate/K₂CO₃ in acetone gave methyl 5-iodo-2-methoxybenzoate 17, which was reacted with SO₂Cl₂ to result in complete chlorination of the aromatic ring at the C3 position. The resulting ester 18 was then hydrolyzed in aqueous NaOH to afford the key intermediate 19 in 95% yield. Coupling of 19 with hydroxymethylpolystyrene resin (Wang resin) under standard DCC coupling conditions (DCC, 3 equiv; HOBt, 3 equiv in DMF) afforded the desired support-bound ester 20.33 It was found that a double-coupling protocol consistently provided much higher loading of the acid onto the solid support [based on elemental analysis (I or Cl) of the modified resin]. Palladium-catalyzed Sonogashira reaction of 20 with 5-hexynoate afforded the resin-bound alkyne **21**. ^{14,15} The IR spectrum (KBr) of the derivatized resin 21 revealed a new absorption at 2232 cm⁻¹ (internal carbon-carbon triple bond), indicating that the desired Sonogashira coupling had occurred. The resin-bound alkyne 21 was then treated with Bu₃SnH in the presence of Pd(PPh₃)₄ to afford 22. The overall efficiency of Sonogashira and hydrostannation reactions was determined to be 86% on the basis of the yield of 23 obtained by basic cleavage (transesterfication, Et₃N/MeOH 15:85, v/v). The Stille coupling of 22 with methyl 3-methyl-5-iodo-2-methoxybenzoate (10) or methyl 5-iodo-2-methoxybenzoate (17) using Pd₂(dba)₃, AsPh₃, and CsF in DMF afforded 24 and 25.16 Cleavage of 24 and 25 with sodium methoxide in methanol and THF (40:60, v/v) afforded ADAMs 26 and 27, respectively. ¹⁴ ¹H NMR analysis of crude products cleaved from the resin showed purities ranging from 75% to 85%. Both 26 and 27 were further purified by silica gel flash chromatography (EtOAc/hexanes 1:3, v/v).

To evaluate the anti-HIV activities of ADAMs **15**, **26**, and **27**, all three compounds were tested for inhibition of the cytopathic effect of HIV- $1_{\rm RF}$ in CEM-SS lymphocytes, and the resulting EC₅₀ values are displayed in Table 1. In addition, these compounds were tested for inhibition of HIV-1 reverse transcriptase with poly(rC)-oligo(dG) as the template primer, and the IC₅₀ values are also reported in Table 1. The cytotoxicities of the compounds in uninfected CEM-SS cells were also investigated, and the 50% cytotoxic concentrations (CC₅₀ values) are also shown.

It is clear from examination of the numbers in Table 1 that there is not an exact correlation between the IC_{50} values for inhibition of HIV-1 reverse transcriptase and inhibition of the cytopathic effect of the virus. For example, for compound 15, the IC_{50} of 0.804 μM for

enzyme inhibition is less than the EC₅₀ of 2.26 μ M for inhibition of the viral cytopathic effect, whereas for ADAMs **26** and **27**, the IC_{50} values of 0.516 and 0.587 μ M are significantly greater than the EC₅₀ values of 0.020 and 0.080 μ M. However, our prior examination of the ADAMs in cells containing mutant HIV-1 reverse transcriptase leave little doubt that the cellular effects of these compounds as inhibitors of cell killing by the virus are in fact due to inhibition of reverse transcriptase.³⁻⁵ Moreover, the lack of correlation of the enzyme IC₅₀ values determined with poly(rC)·oligo(dT) as the template primer and the EC₅₀ values is not unusual for the non-nucleoside reverse transcriptase inhibitors.^{34–37} The discrepancy may simply reflect the difference between the enzyme assay, in which a synthetic template/primer is used, and the cellular system, in which the native viral RNA serves as the template/primer.³⁶

Examination of the antiviral activities of compounds **26**, **27**, and the previously synthesized compound **28**⁴ allows an assignment of the effect of variation of the R group as shown in structures **26** and **27**. The relative anti-HIV potencies are $Cl > CH_3 > H$.

In conclusion, we have devised a new synthesis of ADAMs that allows the incorporation of nonidentical aryl groups in a stereochemically defined fashion. The synthesis can be performed both in solution and on a solid support. The greater flexibility of the present route should allow the future synthesis of non-nucleoside reverse transcriptase inhibitors in the ADAM series having optimized biological activities.

Experimental Section

General. Melting points were determined in capillary tubes on a Mel-Temp apparatus and are uncorrected. The proton nuclear magnetic resonance spectra (1 H NMR) of compounds **18**, **19**, **26**, and **27** were recorded on a 500 MHz spectrometer, and the 1 H NMR spectra of the remaining compounds were determined at 300 MHz. The chemical shift values are expressed in ppm (parts per million) relative to tetramethylsilane as internal standard. The indicated $J_{\rm SnH}$ must be considered as an approximate mean value of $J_{\rm C}^{(117}{\rm SnH})$ and $J_{\rm C}^{(119}{\rm SnH})$. The $^{13}{\rm C}$ NMR spectra were recorded at 75 MHz. As a rule, mass spectra of alkenyltributylstannanes are characterized by the presence of an important peak (often the

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base peak) at $\mathrm{M^+}-57$, which corresponds to the loss of an n-butyl fragment. The $\mathrm{M^+}$ peak was, in the case of **23**, not detected. Microanalyses were performed at the Purdue Microanalysis Laboratory, and all values were within 0.4% of the calculated compositions. Silica gel used for column chromatography was 230-400 mesh.

Methyl 2-Hydroxy-5-iodo-3-methylbenzoate TMSCHN₂ (9 mL of 2 M solution in hexanes) was added to a stirred mixture of 3-methyl salicylic acid (8) (2.28 g, 15 mmol) in methanol (20 mL) and benzene (20 mL) at room temperature. The mixture was stirred for 2 h and concentrated to afford the corresponding methyl ester³⁹ quantitatively. The methyl ester (2.49 g, 15 mmol) was dissolved in methanol (40 mL). Sodium iodide (2.25 g, 15 mmol) and sodium hydroxide (0.60 g, 15 mmol) were added, and the solution was cooled to 0 °C. Aqueous sodium hypochlorite (22.0 g, 5.25% NaOCl) was added dropwise. The brown mixture was stirred for 3 h at 0-3 °C and then treated with 10% aqueous sodium thiosulfate. The pH of the mixture was adjusted to 6−7 using 1 N HCl. Ether (100 mL) was added, and the layers were separated. The ether layer was washed with brine and dried over anhydrous sodium sulfate. After the ether was evaporated, the crude solid was crystallized from a mixture of acetone and hexanes (1:1, v/v) to afford **9** as a yellow solid (4.0 g, 91.3%): mp 82-84 °C; ¹H NMR (CDCl₃) δ 10.94 (s, 1 H), 7.95 (d, J = 2.1 Hz, 1 H), 7.56 (d, J = 1.53 Hz, 1 H), 3.92 (s, 3 H), 2.19 (s, 3 H); 13 C NMR $(CDCl_3)$ δ 169.67, 159.69, 144.33, 135.61, 129.44, 113.56, 79.59, 52.43, 15.24; CIMS m/z 293 (MH⁺).

Methyl 5-Iodo-2-methoxy-3-methyl-benzoate (10). Compound 9 (3.0 g, 10.27 mmol) was dissolved in acetone (50 mL). Anhydrous K₂CO₃ (4.2 g, 30.47 mmol) and dimethyl sulfate (3.88 mL, 41.08 mmol) were added. The reaction mixture was vigorously stirred under reflux for 24 h and then filtered, and the inorganic salts were washed with acetone (45 mL). The solvent was evaporated, and the yellow residue was washed with water (3 \times 100 mL) to remove excess dimethyl sulfate and crystallized from acetone to afford 10 as white crystals (3.1 g, 99%): mp 55-57 °C; ¹H NMR (CDCl₃) δ 7.93 (d, J =2.28 Hz, 1 H), 7.65 (d, J = 2.20 Hz, 1 H), 3.90 (s, 3 H), 3.80 (s, 3 H), 2.27 (s, 3 H); $^{13}\mathrm{C}$ NMR (CDCl₃) δ 165.21, 158.26, 143.43, 137.55, 135.34, 126.32, 86.54, 61.49, 52.27, 15.60; EIMS m/z 306 (M⁺); HRMS calcd for C₁₀H₁₂IO₃ (MH⁺) 306.9831, found 306.9828. Anal. Calcd for C₁₀H₁₁IO₃: C, 39.34; H, 3.61. Found: C, 39.01; H, 3.50.

Methyl 5-(Tri-n-butylstannyl)-2-methoxy-3-methylbenzoate (11). Compound 10 (612 mg, 2.0 mmol) was dissolved in triethylamine (20 mL). Palladium(II) acetate (34 mg, 0.148 mmol) and tetrakis(triphenylphosphine)palladium(0) (86 mg, 0.074 mmol) were added, followed by bis(tri-n-butyltin) (1.75 mL, 3.45 mmol). The mixture was heated at 95-100 °C for 3 h. The black reaction mixture was filtered, and the black solid was washed with triethylamine (3 \times 10 mL). The filtrate was evaporated under reduced pressure. The oily residue was purified by flash chromatography on silica gel (120 g, column 5 cm imes 31 cm), eluting with a mixture of ethyl acetate and hexanes (1:5, v/v), to afford a colorless oil (600 mg, 64%): 1H NMR (CDCl₃) δ 7.67 (s, 1 H), 7.39 (s, 1 H), 3.92 (s, 3 H), 3.82 (s, 3 H), 2.31 (s, 3 H), 1.62-1.27 (m, 16 H), 1.05 (t, J = 8.11Hz, 2 H), 0.92 (t, J = 7.30 Hz, 3 H), 0.89 (t, J = 7.24 Hz, 6 H); ¹³C NMR (CDCl₃) δ 167.44, 158.17, 143.01, 136.56, 131.72, 124.01, 118.33, 61.29, 51.99, 29.02, 28.90, 27.19, 26.51, 16.29, 15.88, 13.52, 13.45, 9.52; ESI m/z 469 (MH⁺); HRMS calcd for C₂₂H₃₉SnO₃ (MH⁺) 469.1915, found 469.1934. Anal. Calcd for C₂₂H₃₈SnO₃: C, 56.41; H, 8.12. Found: C, 56.22; H, 7.90.

N-Methyl-6-[4-methoxy-5-(methoxycarbonyl)-3-methylphenyl]hex-5-ynoamide (12). 5-Hexynoic acid (1.0 g, 8.92 mmol) was dissolved in dry THF (15 mL). Oxalyl chloride (1.17 mL, 13.38 mmol) was added, followed by a catalytic amount of DMF (1 drop). The mixture was stirred for 45 min at room temperature. Excess solvent and oxalyl chloride were removed

under reduced pressure. The resulting residue was dissolved in dry THF (10 mL) and cooled to 0 °C. Methylamine (10 mL of 2 M methylamine in THF) and triethylamine (0.9 g, 8.92 mmol) were added. The mixture was stirred for 2 h and then washed with water (2 \times 5 mL). The organic solvent was evaporated, and the crude residue was further purified by flash chromatography on silica gel (40 g, column 1.5 cm \times 31 cm; eluent, ethyl acetate) to give N-methyl-5-hexynoamide40 as a light yellow oil (0.81 g, 73%): 1 H NMR (CDČl₃) δ 5.72 (br, 1 \dot{H}), 2.79 (d, J = 4.47 \dot{H} z, 3 H), 2.30 (t, J = 7.34 Hz, 2 H), 2.23 (td, J = 6.88 and 2.24 Hz, 2 H), 1.95 (t, J = 2.23 Hz, 1 H), 1.84 (m, 2 H); ¹³C NMR (CDCl₃) δ 172.76, 83.42, 68.99, 34.80, 26.15, 24.04, 17.70. The amide (375 mg, 3.0 mmol), methyl 5-iodo-2-methoxy-3-methylbenzoate 4 (875 mg, 2.73 mmol), Pd(Ph₃P)₂(OAc)₂ (145 mg, 0.194 mmol), copper(I) iodide (104 mg, 0.546 mmol), and triethylamine (791 mg, 7.84 mmol) were stirred in ethyl acetate (25 mL) at room temperature for 14 h under argon. The mixture was filtered through a pad of Celite, and the Celite was washed with a mixture of ethyl acetate and hexanes (20 mL, 1:1, v/v). The combined filtrates were evaporated, and the brown residue was purified by flash chromatography (silica gel 45 g, column 1.5 cm \times 31 cm), eluting with ethyl acetate/hexanes (1:1, v/v), to afford a colorless oil (800 mg, 80%): 1 H NMR (CDCl₃) δ 7.65 (d, J = 2.04 Hz, 1 H), 7.35 (d, J = 2.04 Hz, 1 H), 5.63 (br, 1 H), 3.92 (s, 3 H), 3.80 (s, 3 H), 2.81 (d, J = 4.92 Hz, 3 H), 2.44 (t, J =6.79 Hz, 2 H), 2.34 (t, J = 7.41 Hz, 2 H), 2.26 (s, 3 H), 1.91 (m, 1.91 m)2 H); ^{13}C NMR (CDCl3) δ 172.82, 166.14, 157.78, 137.71, 132.85, 132.23, 131.87, 128.48, 128.33, 124.40, 119.01, 88.96, 61.46, 52.13, 35.05, 26.15, 24.35, 18.72, 15.76; CIMS m/z 304 (MH+); HRMS calcd for C₁₇H₂₂NO₄ (MH+) 304.1549, found 304.1534. Anal. Calcd for C₁₇H₂₁NO₄: C, 67.33; H, 6.93. Found: C, 67.12; H, 6.87.

Methyl 5-[(1*E*)-5-(*N*-Methylcarbamoyl)-1-(tributylstannyl)pent-1-enyl]-2-methoxy-3-methylbenzoate (13). Bu₃-SnH (1.3 mL, 4.85 mmol) was added dropwise to a stirred solution of alkyne 12 (1.0 g, 3.3 mmol) and (Ph₃P)₄Pd (109.4 mg, 0.095 mmol) in dry THF (25 mL) at room temperature. The reaction mixture was stirred for 40 min under argon. The solvent was evaporated, and the brown residue was purified by flash chromatography on silica gel (100 g, EtOAc/hexanes 2:3, v/v) to afford a light brown oil (1.70 g, 86.7%): ¹H NMR (CDCl₃) δ 7.21 (d, J = 2.28 Hz, 1 H), 6.92 (d, J = 2.07 Hz, 1 H), 5.75 (t, J = 6.96 Hz, 1 H, ${}^3J_{\rm SnH}$ = 33 Hz), 5.40 (br, 1 H), 3.95 (s, 3 H), 3.84 (s, 3 H), 2.74 (d, J = 4.89 Hz, 3 H), 2.32 (s, 3 H), 2.11 (t, J = 7.39 Hz, 2 H), 2.08 (m, 2 H), 1.73 (m, 2 H), 1.23-1.51 (m, 18 H), 0.90 (t, J = 7.0 Hz, 9 H); 13 C NMR (CDCl₃) δ 173.30, 166.99, 155.58, 144.86, 141.48, 140.13, 133.60, 132.16, 127.25, 123.84, 61.37, 51.97, 35.82, 29.26, 28.84, 27.14, 26.06, 25.55, 16.00, 13.53, 9.85; ESI m/z 594/596 (MH⁺). Anal. Calcd for C₂₉H₄₉NSnO₄.CH₃COOC₂H₅: C, 58.15; H, 8.37. Found: C, 57.86; H, 8.16.

Methyl 5-[(1*E*)-1-Iodo-5-(*N*-methylcarbamoyl)pent-1enyl]-2-methoxy-3-methylbenzoate (14). The vinyl tributylstannane 13 (594 mg, 1.0 mmol) was dissolved in dry CH₂Cl₂ (20 mL). Finely divided I₂ (305 mg, 1.2 mmol) was added, and the mixture was stirred vigorously at room temperature for 20 min. Saturated aqueous Na₂S₂O₃ (15 mL) was added, the phases were separated, and the aqueous phase was extracted with CH₂Cl₂ (3 ×10 mL). The combined organic extracts were dried (Na₂SO₄) and concentrated. The residue was purified by flash chromatography on silica gel (30 g, EtOAc as eluent) to afford a brown oil (310 mg, 71.9%): 1 H NMR (CDCl₃) δ 7.51 (d, J = 2.31 Hz, 1 H), 7.23 (d, J = 2.01 Hz, 1 H), 6.43 (t, J =7.60 Hz, 1 H), 5.50 (br, 1 H), 3.91 (s, 3 H), 3.82 (s, 3 H), 2.71 (d, J = 4.87 Hz, 3 H), 2.29 (s, 3 H), 2.08 (t, J = 7.48 Hz, 2 H), 2.10 (dt, J = 7.34 and 7.33 Hz, 2 H), 1.71 (m, 2 H); ¹³C NMR (CDCl₃) δ 172.75, 166.32, 157.78, 142.98, 136.81, 135.08, 132.85, 129.12, 124.08, 93.59, 61.43, 52.22, 35.37, 31.36, 26.12, 24.77, 16.03; CIMS m/z 432 (MH+); HRMS calcd for C₁₇H₂₂-NIO₄ 432.0672, found 432.0671. Anal. Calcd for C₁₇H₂₂NIO₄: C, 47.33; H, 5.10; N, 3.25; I, 29.47. Found: C, 47.52; H, 5.22; N, 3.21; I, 29.36.

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Methyl 3,3'-Dimethyl-4,4'-dimethoxy-5,5'-bis(methoxycarbonyl)-6,6-diphenylhexenoamide (15). Method A. Under an atmosphere of argon, a solution of iodo ester 10 (60 mg, 0.2 mmol) in dry THF (5 mL) and a solution of AsPh₃ (24.5 mg, 0.08 mmol) in dry THF (5 mL) were added in turn to a two-necked flask charged with Pd₂(dba)₃ (18.3 mg, 0.02 mmol) and CsF (67 mg, 0.44 mmol). The organostannane 13 (120 mg, 0.2 mmol in 5 mL of dry THF) was then added by syringe, and the flask was placed in an oil bath (70-80 °C) and stirred for 14 h under argon. The reaction mixture was cooled to room temperature, diluted with Et₂O, and filtered through a pad of Celite. The Celite was washed throughly with Et₂O, and the combined filtrates were concentrated by rotary evaporation. The product was purified by flash chromatography on silica gel (30 g), eluting with ethyl acetate, to afford the desired ĀDAM-amide **15** (71 mg, 74%): 1 H NMR (CDCl₃) δ 7.49 (d, J= 2.12 Hz, 1 H), 7.44 (d, J = 1.75 Hz, 1 H), 7.13 (m, 2 H), 6.01(t, J = 7.38 Hz, 1 H), 3.96 (s, 3 H), 3.94 (s, 3 H), 3.91 (s, 3 H), 3.79 (s, 3 H), 2.78 (d, J = 5.02 Hz, 3 H), 2.35 (s, 3 H), 2.28 (s, 3 H), 2.21 (t, J = 7.55 Hz, 2 H), 2.16 (dt, J = 7.60 and 6.11 Hz, 2 H), 1.76-1.85 (m, 2 H); CIMS m/z 484 (MH)+; HRMS calcd for C₂₇H₃₃NO₇ 483.2257, found 483.2263. Anal. Calcd for C₂₇H₃₃NO₇: C, 67.08; H, 6.83. Found: C, 66.97; H, 6.79.

Method B. Under an atmosphere of argon, a solution of vinyl iodide 14 (54.2 mg, 0.13 mmol) in dry THF (3 mL) and a solution of AsPh₃ (15.9 mg, 0.05 mmol) in dry THF (3 mL) were added in turn to a two-necked flask charged with Pd₂(dba)₃ (11.9 mg, 0.01 mmol) and CsF (43.6 mg, 0.29 mmol). The organostannane 11 (61.9 mg, 0.13 mmol) in dry THF (4 mL) was then added by syringe, and the flask was placed in an oil bath (70-80 °C) and stirred overnight under argon. The reaction mixture was cooled to room temperature, diluted with Et₂O, and filtered through a pad of Celite. The Celite was washed thoroughly with Et₂O, and the combined filtrates were concentrated by rotary evaporation. The product was purified by flash chromatography on silica gel (25 g), eluting with ethyl acetate, to afford the desired ADAM-amide 15 (47 mg, 84%).

Methyl 3-Chloro-5-iodo-2-methoxybenzoate (18). To a solution of methyl 5-iodo-2-methoxybenzoate 1741 (2.92 g, 10 mmol) in CH₂Cl₂ (10 mL) was gradually added SO₂Cl₂ (5.4 g, 40 mmol). The reaction mixture was heated at 50 °C for 14 h and then cooled to room temperature. It was poured into a large amount of water (200 mL) and extracted with CH₂Cl₂ (3 × 40 mL). The CH₂Cl₂ extracts were combined, dried over Na₂SO₄, and evaporated in vacuo to afford a white solid (3.20 g, 98%). A small amount of the sample was further purified by flash chromatography on silica gel (10 g, hexanes as eluent): mp 41–43 °C; ¹H NMR (CDCl₃) δ 7.97 (d, J = 2.12Hz, 1 H), 7.84 (d, J = 2.08 Hz, 1 H), 3.90 (s, 6 H); 13 C NMR (CDCl₃) δ 164.25, 156.14, 142.05, 138.42, 130.79, 128.33, 86.15, 62.00, 52.63. Anal. Calcd for C₉H₈IClO₃·0.6 H₂O: C, 32.05; H, 2.75. Found: C, 31.90; H, 2.35.

3-Chloro-5-iodo-2-methoxybenzoic Acid (19). A 1 N NaOH solution (10 mL) was added to a MeOH solution (8 mL) containing benzoate 18 (0.9 g, 2.76 mmol). The mixture was heated at 60 °C for 3 h and then acidified with concentrated HCl solution (the pH was adjusted to 1-2). The white precipitate was filtered and recrystallized from acetone to afford compound **19** (0.82 g, 95%): mp 189-191 °C (dec); ¹H NMR (CD₃COCD₃) δ 8.04 (d, J = 2.24 Hz, 1 H), 7.98 (d, J =2.13 Hz, 1 H), 3.90 (s, 3 H); 13 C NMR (CD₃COCD₃) δ 169.14, 160.87, 146.72, 143.67, 134.32, 91.00, 66.60; CIMS m/z 313. Anal. Calcd for C₈H₆IClO₃: C, 30.72; H, 1.92; Cl, 11.36; I, 40.64. Found: C, 30.42; H, 1.68; Cl, 11.08; I, 40.33.

Esterification of Wang Resin with 3-Chloro-5-iodo-2methoxybenzoic Acid (19). Solid DCC (630 mg, 3.05 mmol) and HOBt (621 mg, 3.05 mmol) were added to a suspension of Wang resin (1.21 mmol –OH, 1 g) in a mixture of dry DMF and pyridine (30 mL, 9:1 v/v). 3-Chloro-5-iodo-2-methoxybenzoic acid (19) (406 mg, 1.30 mmol) was added, and the mixture was gently stirred at room temperature for 24 h. The suspension was filtered, and the resin was washed with pyridine (25 mL) and DMF, CH₂Cl₂, and MeOH (3 × 25 mL each) and then dried under reduced pressure over P2O5. Elemental analysis of the resin **20** (first cycle): I, 3.21; Cl, 0.93. This procedure was repeated again on this resin to afford 20. IR (KBr) 1726.3 cm⁻¹, 1492.2 cm⁻¹. Elemental analysis of the resin **20** (second cycle): I, 12.70; Cl, 3.80.

Pd(0)-Catalyzed Sonogashira Coupling to Afford Resin 21. The resin 20 (950 mg) was suspended in a mixture of dry DMF and THF (30 mL, 5:1, v/v). Methyl 5-hexynoate (434 mg, 3.44 mmol), Pd(OAc)₂ (21 mg, 0.092 mmol), PPh₃ (60.3 mg, 0.23 mmol), CuI (44 mg, 0.23 mmol), and triethylamine (337 mg, 3.34 mmol) were sequentially added to a round-bottomed reaction flask containing the resin 20. The reaction flask was then stoppered, and the reaction mixture was stirred overnight under argon. The solvent and excess reactants were then removed by filtration, and the remaining light brown solid support was washed with DMF, THF, MeOH, and CH2Cl2 (3 × 25 mL of each) and dried in vacuo. IR (KBr) 2232 cm⁻¹; 1731.4 cm⁻¹.

Hydrostannation of Resin-Bound Alkyne 21 to Yield **Resin 22.** Pd(PPh₃)₄ (35 mg, 0.03 mmol) was added to a degassed suspension of polymer-bound alkyne 21 (0.83 g, ~1 mmol) in a mixture of dry THF and DMF (30 mL, 2:1 v/v). The mixture was stirred for 5 min, and Bu₃SnH (0.41 mL, 1.5 mmol) was then slowly added via syringe over 10 min. The reaction mixture was allowed to stir at room temperature for 4 h under argon, transferred to a glass filter, and thoroughly washed with CH_2Cl_2 , MeOH, DMF, and then CH_2Cl_2 (3 × 25 mL each) to afford the desired solid support 22.

Cleavage of Compound 23 from Solid Support 22. The cleavage cocktail was prepared by degassing a solution of MeOH (17 mL), THF (5 mL), and Et₃N (3.0 mL) with argon. This solution was added to the dried resin 22 (100 mg), and the mixture was heated at 50 °C under argon for 16 h at room temperature. The resin was filtered and washed with MeOH (2 mL). The resin was subjected to this procedure a total of six times. The combined filtrates were evaporated to afford compound **23** (64 mg): ¹H NMR (CDCl₃) δ 7.16 (d, J = 2.17Hz, $\hat{1}$ H), 7.02 (d, J = 2.18 Hz, 1 H), 5.68 (t, J = 6.95 Hz, 1 H, ${}^{3}J_{SnH} = 31.1 \text{ Hz}$), 3.86 (s, 3 H), 3.85 (s, 3 H), 3.56 (s, 3 H), 2.19 (t, J = 7.55 Hz, 2 H), 1.99 (dt, J = 7.04 and 7.33 Hz, 2 H), 1.61 (m, 2 H), 1.14–1.45 (m, 18 H), 0.90 (t, J = 7.0 Hz, 9 H). Anal. Calcd for C₂₈H₄₅ClSnO₅: C, 54.61; H, 7.37; Cl, 5.76. Found: C, 54.43; H, 7.31; Cl, 5.68.

Pd(0)-Catalyzed Stille Coupling to Afford Resins 24 and 25. A solution of AsPh3 (24.5 mg, 0.08 mmol) in THF (5 mL) was added to a degassed suspension of polymer-bound vinylstannane 22 (400 mg) in anhydrous DMF (15 mL) containing Pd₂(dba)₃ (19.6 mg, 0.0214 mmol) and CsF (67 mg, 0.44 mmol). After 5 min, aryl iodide **10** (391 mg, 1.28 mmol) in anhydrous DMF (10 mL) was added, and the reaction mixture was heated at 80 °C overnight under argon. The reaction mixture was then transferred to a fritted syringe and thoroughly washed with DMF, CH₂Cl₂, EtOH, and CH₂Cl₂ (3 × 25 mL each) to afford solid support 24, which was dried under reduced pressure over P_2O_5 . A similar procedure was employed to prepare solid support 25 via Stille coupling of aryl iodide 17 with resin-bound vinylstannane 22.

Cleavage of ADAMs 26 and 27 from Resins 24 and 25. The appropriate resin 24 or 25 (200 mg) was added to a degassed MeOH/THF solution (60 mL, 1:2 v/v) containing NaOMe (167 mg, 3 mmol). The reaction mixture was heated under reflux for 20 h. The resin was filtered and washed with MeOH (10 mL), and the filtrate was evaporated under reduced pressure to afford crude ADAM 26 (73 mg) or 27 (92 mg) as a light yellow liquid. Both compounds were further purified by flash chromatography on silica gel (EtOAc/hexanes 1:3, v/v).

Methyl (5Z)-6-[3-Chloro-4-methoxy-5-(methoxycarbonyl)phenyl]-6-[4-methoxy-5-(methoxycarbonyl)-3-methylphenyl]hex-5-enoate (26). Yield 54.7 mg; ¹H NMR (CDCl₃) δ 7.62 (d, J = 2.40 Hz, 1 H), 7.56 (d, $J = \check{2}.06$ Hz, 1 H), 7.37 (d, J = 2.30 Hz, 1 H), 7.34 (d, J = 2.20 Hz, 1 H), 5.93 (t, J =7.46 Hz, 1 H), 3.92 (s, 3 H), 3.87 (s, 3 H), 3.83 (s, 3 H), 3.73 (s, 3 H), 3.57 (s, 3 H), 2.24 (t, J = 7.32 Hz, 2 H), 2.19 (s, 3 H),

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2.06 (dt, J= 7.58 and 7.43 Hz, 2 H), 1.68 (m, 2 H). Anal. Calcd for $C_{26}H_{29}ClO_8$: C, 61.84; H, 5.79; Cl, 7.02. Found: C, 62.13; H, 5.77; Cl, 6.83.

Methyl (5*Z*)-6-[3-Chloro-4-methoxy-5-(methoxycarbonyl)phenyl]-6-[4-methoxy-3-(methoxycarbonyl)phenyl]-hex-5-enoate (27). Yield 79.0 mg; 1 H NMR (CDCl₃) δ 7.63 (d, J = 2.38 Hz, 1 H), 7.52 (d, J = 2.46 Hz, 1 H), 7.30 (d, J = 2.12 Hz, 1 H), 7.15 (dd, J = 8.59 and 2.34 Hz, 1 H), 6.84 (d, J = 8.79 Hz, 1 H), 5.97 (t, J = 7.47 Hz, 1 H), 3.96 (s, 3 H), 3.92 (s, 3 H), 3.89 (s, 3 H), 3.86 (s, 3 H), 3.61 (s, 3 H), 2.29 (t, J = 7.07 Hz, 2 H), 2.12 (dt, J = 7.43 and 7.23 Hz, 2 H), 1.79 (m, 2 H); EIMS m/z 490 (M⁺). Anal. Calcd for C₂₅H₂₇ClO₈: C, 61.10; H, 5.50; Cl; 7.24. Found: C, 61.28; H, 5.49; Cl, 7.12.

HIV Cytoprotection Assay. Anti-HIV screening of test compounds was performed as previously described^{42,43} with minor changes. Briefly, the assays involve the killing of T4 lymphoid cells (CEM-SS cell line) by HIV-1 and inhibition of cell killing by active compounds. Experimental antiviral compounds were diluted as appropriate and added to a 96-well microtiter plate. Cells were infected in the well at a multiplicity of infection (MOI) of approximately 0.2, and viability was assayed at 6 days after infection by reduction of the CellTiter 96 Reagent (Promega, Madison, WI). This reagent contains the tetrazolium compound 3-(4,5-dimethylthiazol-2-yl)-5-(carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H

tetrazolium, inner salt (MTS) and the electron-coupling agent phenazine ethosulfate in a colorless stable solution, which upon reduction by viable cells forms a colored solution with absorbance at 490 nm. Data are presented as the percent control of MTS values for the uninfected, drug-free control. EC₅₀ values reflect the drug concentration that provides 50% protection from the cytopathic effect of HIV-1 in infected cultures, while the CC₅₀ reflects the concentration of drug that causes 50% cell death in the uninfected cultures. MTS-based results were confirmed by measurement of cell-free supernatant RT and p24 levels. All MTS cytoprotection data were derived from triplicate tests on each plate, with two separate sister plates. Thus, the EC₅₀ value from each plate represents the average of triplicates, and the two EC₅₀ values from sister plates were averaged. The variation from the mean averaged less than 10%.

Assay for Inhibition of HIV-1 RT. The effects of the compounds on the HIV-1 RT enzyme were performed using recombinant purified p66/51 RT (a kind gift of S. Hughes, NCI-FCRDC). Inhibition of reverse transcription was measured by the level of incorporation of [32P]GTP into the poly(rC)·oligo-(dG) (rCdG) homopolymer template primer system.³

Acknowledgment. This investigation was made possible by grant RO1-AI-43637, awarded by the National Institutes of Health, DHHS. We thank David Allen for the creation of the cover art.

JO0100291

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